

Progesterone and insulin - like growth factor 1 in serum of Shami goats during the early breeding season

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SUMMARY

The study aimed to determine the level and characterize the plasma IGF-1 profiles with relationship to progesterone profiles during the early breeding season in Shami goats. 10 multiparous- non pregnant Shami does, kept indoors at night and outside for most of the day were offered diets based on barley and wheat straws with free access to natural grazing, water and minerals ad libitum. Daily prefeeding- blood samples (10 ml) for 46 days were collected in heparinized tubes by jugular venipuncture and centrifuged at 3000 rpm for 20 min. Serum was harvested and stored at -20°C until assayed for hormonal analysis using radioimmunoassay technique. Data were analyzed as a repeated measure design using PROC GLIIMMIX Procedures in SAS 9.4. Differences in least square means for IGF-1 and progesterone were tested using Fisher's protected least significant differences ($p \leq 0.05$). Results showed only 20% of Shami goats showed ovarian activity based on P4 serum concentration during the early breeding period. No significant effect for the studied groups (cyclic vs. non cyclic) was observed, but significant effects ($p < 0.03$) for the seasonal days and interaction with the animal groups ($p < 0.02$) on the concentration of IGF-1 were observed. A negative and significant relationship ($r = -0.39$, $p < 0.01$) were obtained between levels of IGF-1 and progesterone and the peak of IGF-1 preceded the p4 peak by 12 days in cycling does. Determination the concentrations of these two hormones can help in assessing the reproductive status of Shami goats and facilitate the ability to select females most suited higher reproductive efficiency.

Keywords: Shami goats, breeding season, cyclicity, IGF-1, progesterone.

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INTRODUCTION

Reproductive seasonality limits the reproductive efficiency in goat production systems (Shelton, 1991). The yearly reproductive cycle of the goat is primarily controlled by an endogenous reproductive rhythm, which is synchronized by the day length (i.e. photoperiod) (Delgadillo et al., 2004). The duration of anestrus can also be modulated by nutritional and socio-sexual factors in sheep and goats (Martin et al., 1999; Forcada and Abecia, 2006).

Better understanding of the relationship between nutrition and ovarian function is a fundamental cue to optimizing the reproductive parameters. In goats, as with other ruminants, the nutritional effect on ovulation and follicular recruitment was mainly explained by short period influence of energetic status on gonadotropin secretion (Mani et al., 1996). Rondina et al. (2005) indicated that opposite dietary intakes for a medium period can regulate the early onset of follicle growth of local goats. Undernutrition for short (Mani et al., 1992; 1996), medium (Tanaka et al., 2004) or long period (Paula et al., 2005) has been extensively reported to exert a negative influence on ovarian activity in goats and level of progesterone secretion.

Some investigators have reported the concentrations of plasma progesterone during the estrous cycle in goats for different breeds (Bauernfeind and Holtz, 1991; El-Hommosy et al., 1991; Pathiraja et al., 1991), while Mavrogenis (1988), Shalaby et al. (2002), Khadiga et al., (2005) and Zarkawi and Soukouti (2001) studied changes in the plasma progesterone concentration during different stages of the estrous cycle in Shami goats.

Many studies have highlighted the role of the insulin-like growth factor I (IGF-I) system in the control of follicular growth (Giudice, 1992; Armstrong and Webb, 1997; Armstrong et al., 1998; Woad et al., 2000). The responsiveness of ovarian follicles to IGF-I has been demonstrated in numerous studies. When added during the *in vitro* culture of preantral follicles, IGF-I has been shown to stimulate follicular growth in various species (rat: Zhou et al., 2003; cow: Gutierrez et al., 2000; human: Louhio et al., 2000). In addition, IGF-I increases follicular diameter, DNA content, promote functional integrity and stimulated steroidogenesis during rat preantral follicle culture (Demeestere et al., 2004). However, much less is known about the involvement of IGF-I in the regulation of goat preantral follicular development.

It is reported that plasma concentration of IGF-1 increases during estrus in ruminants (Spicer and Zavy, 1992; Hashizume et al., 2000) and thought to function primarily as a circulating hormone secreted by the liver in response to GH in domestic animals (Hossner, et al., 1997), in the ovine ovary (Perks et al., 1995), in oviduct of bovine (Schmidt et al., 1994) and

ovine (Stevenson and Wathes, 1996), in ruminants and rat uterus (Wathes et al., 1998) and from the uterus in goats (Nonaka et al., 2003). It has been speculated that most of IGF-1 in blood during estrus in ruminants might be derived from the uterus (Wathes et al., 1998; Stevenson et al., 1994; Geisert et al., 1991; Ko et al., 1991).

Shami (Damascus) goat is a native breed to Syria and requires an improvement management and feeding environment to express its all genetic potential (Mavrogenis et al., 2006). It is a highly adaptive and prolific animal that is recognized as the ultimate goat in either dual or single purpose production systems. It can be used in pure bred or cross-bred milk or meat production systems, surpassing the production figures of equivalent animals in intensive, semi-intensive or extensive systems (Constantinou, 1989). Shami goats are considered as the most important goat breed in the Arab and West Asian countries, due to its high milk yield (450-850 kg/260 days) and meat production potential (1.8 litter size) (Abdullah et al., 2012). It is a seasonal breed, and the occurrence of the first seasonal estrus is somewhat irregular. Does exhibit a short (August–October) and sometimes long breeding season (July–November), rarely demonstrating estrus during winter (December and January) in Syria (Al Khouri, 1996). The main breeding season starts in late August and extends through mid-December in Cyprus (Abdullah et al., 2012).

Certain reproductive parameters in Damascus does have been studied such as oestrous cycle (Zarkawi and Soukouti 2001), duration of pregnancy (Zarkawi and Al-Masri 2002), early pregnancy diagnosis and monitoring corpora lutea (Zarkawi et al., 2003) and the response to some veterinary agents (Zarkawi, 2007).

The hypothesis of the present research is that alteration in the IGF-1 level might reflex the nutrition status and its influence on reestablishment ovarian cyclicity during the early breeding season in Shami goats and might facilitate the ability of producers to select females most suited for particular nutritional environments and higher reproductive efficiency. This work, therefore, aimed to determine the level and characterize the plasma IGF-1 profiles with relationship to progesterone profiles during the early breeding season in Shami goats.

MATERIAL AND METHODS

Ten multiparous, not pregnant Shami does with a live weight of 55.6 ± 5.06 kg (Mean \pm SEM) reared at Der Al-Hajar Shami Goat Production Research Station, 33 km southeast of Damascus were used. This is considered a dry area with an annual rainfall of about 100 mm occurring mainly in December and January. The does were kept separated from the rest of the herd and kept indoors at night and outside for most of the day.

Indoors, the animals were offered diets based on barley and wheat straw and outdoors, they had free access to natural grazing with major fluctuation and feed availability throughout of the year. Water and minerals were available *ad libitum*. Parasite treatment and estrus detection by introducing a buck two times daily (09:00, 16:00 h) were practiced.

Once a day (9:00 h) and for 46 days (August 11 to September 25), pre feeding- blood samples (10 ml) were collected in heparinized tubes by jugular venipuncture and centrifuged at 3000 rpm for 20 min, serum was harvested and stored at -20°C until assayed. Frozen serum was replaced into a well-three folded box containing 12 kg of frozen gel to keep the samples frozen during the air transportation to the reproductive lab at the Animal Science Research Center (ASRC) at the University of Missouri for hormonal analysis. Insulin-like growth factor 1 (IGF1) and Progesterone were analyzed via double antibody radioimmunoassay previously validated within our laboratory (Morrison et al., 2002; Lalman et al., 2000, Pohler et al., 2015) and inter- and intra-assay CV were < 5%.

The data were analyzed as a repeated measure design outlined by Littell et.al. (1998). The main plot contained the main effect of group (cyclic and Non-cyclic), the subplot contained the effects of day and the interaction of group*day. The denominator of F for main plot was animal identification (group) and for the subplot effects was the residual mean square. The data were analyzed using PROC GLIMMIX Procedure in SAS 9.4. Differences in least square means were tested using Fisher's protected least significant differences (LSD). ($p \leq 0.05$). Correlation was also applied to determine the relationship between the concentration of IGF-1 and progesterone over times in the same animal and between does.

RESULTS

The serum profiles of IGF-1 and progesterone (P4) in Shami goats, measured by RIA during the early breeding season for cyclic and non-cyclic does are presented in Figure 1. Only 2 out of 10 Shami goats showed ovarian activity based on P4 serum concentration during the early breeding period (Fig. 1). In the ANOVA, no significant effect for the studied groups (cyclic vs. non cyclic) was observed, but there were significant effects ($p < 0.03$) for the seasonal days and interaction with the animal groups ($p < 0.02$) on the concentration of IGF-1. Correlation analysis showed a negative and significant relationship ($r = - 0.39$, $p < 0.01$) between levels of IGF-1 and progesterone in Shami does.

No significant changes in the concentration of serum IGF-1 were observed in non-cycling group (Fig. 1). However, does within this group were highly reproducible from day today and the reproducibility of IGF-1 concentration , pooled across lines among days 1 to 10 , 11 to 16, 17 to 34

and 35 to 46 were 1.1, 0.95, and 1.2, respectively. These changes were highly significant ($p < 0.05$) in cycling group and the IGF-1 curve for the two cycling does was similar and the concentration of IGF-1 started to rise from day 11 in animal 7 and day 13 in animal 9 and reached the highest value on day 16 of seasonal days in both does. The mean maximal IGF-1 concentration observed on day 16 was significantly ($p < 0.01$) higher than any following days in cycling animals and any days during the studied period in non-cycling does ($p < 0.05$).

Concentration of P4 was mostly undetectable or lower than 0.3 ng/ml in non-cycling does ($n=8$) (Fig. 1) while it increased significantly ($p < 0.01$) in cycling does ($n=2$) with similar patterns during the study period (Fig. 1).

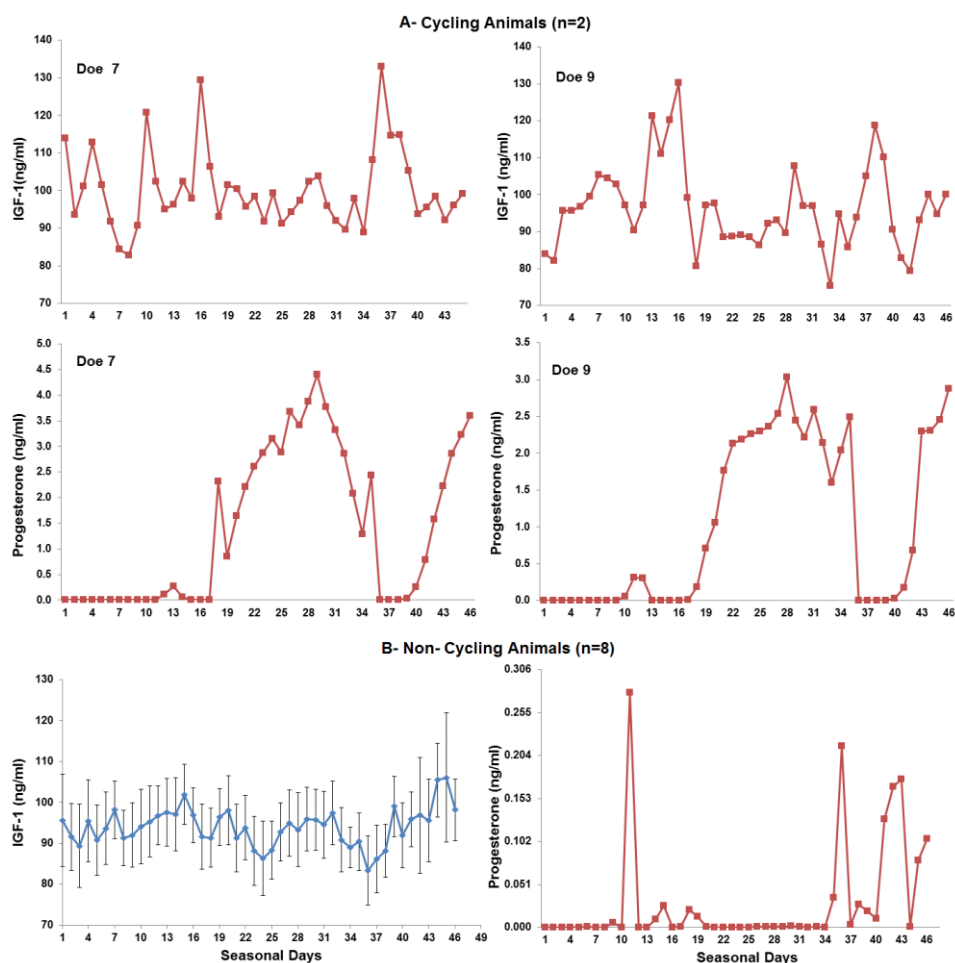


Figure 1. Daily patterns of serum IGF-1 and progesterone in cycling (A, does Nos. 7 and 9) and non-cycling Shame does ($n=8$, B) during the early breeding season (Aug. 11th - Sept. 26th). Hormone concentrations were measured in samples collected on daily basis using RIA technique.

In cycling does, the P4 concentration started to rise on day 19 and peaked up on day 28 (doe 7) and day 27 (doe 9) then gradually decreased to become undetectable on day 36 and showed new sustainable increase after day 41(2nd cycle) during the study period of breeding season. These changes and the increase in p4 concentrations above 1 ng/ml between day 18 and day 35 and day 20 and day 35 of the studied period in does 7 and 9, respectively indicates to the evidence of ovulation and the presence of active corpus luteum. The mean maximal progesterone concentration (3 to 4.45 ng/ml) occurred from day 20 to day 35 of the study period in these cycling does and simultaneous significant reduction in the level of IGF-1 confirms the negative relationship between the secretions of these two hormones.

DISCUSSION

To our knowledge, this is the first investigation to clarify the profile of serum IGF-1 and its relation to P4 profile during the early breeding season in Shami goats.

The overall mean concentration of IGF-1 in Shami goat was 97.64 ± 14.2 ng/ml in cycling does and 93.77 ± 7.11 ng/ml in non- cycling does which is higher than 35 to 80 ng/ml out of breeding season in Boer goats in Croatia (Djuricic et al., 2011) and much less than 150 ng/ml and 250 ng/ml reported by Hashizume et al. (2000) during the luteal and estrus phases, respectively in Japanese goats and also less than 150 to 200 ng/ml during the estrus cycle in sheep (Spicer and Zavy, 1992).

The role of IGF-1 in the reproduction of ruminant has been the subject of intensive studies during the last two decades (Wathes et al., 1998; Tamadon et al., 2010) and its role is very complex, as it occurs on different levels, e.g. at the central hypothalamic level (Todd et al., 2010) and its local level through its influence on ovary activity (Yu et al., 2003; Hunter et al., 2004) and owing to its systemic secretion, mostly by the liver, and simultaneous to local production in different tissues of reproductive tract (Millard et al., 2010). Plasma IGF-1 concentrations increase during the estrus in rats (Carlsson et al., 1989), in Sheep (Spicer and Zavy, 1992; Leeuwenberg et al., 1995) and goats (Hashizume et al., 2000; Nonaka et al., 2003). Hashizume et al. (2000) confirmed that IGF-1 releases from the ovaries during cyclic activity could increase systemic IGF-1 concentrations. Richards et al. (1991) reported that ovariectomy causes a decrease in serum IGF-1 in cattle and the exogenous estradiol causes a significant increase in serum IGF-1 in ovariectomized cows. The consistent and large increase in plasma IGF-1 observed after injection of PGF2 α in goats (Hashizume et al., 2000) and shortly after sponge withdrawal and reaching a maximum about day after the onset of estrus in ewes (Spicer et al., 1993)

suggests that follicular estrogen may regulate IGF-1 secretion in goats and sheep as also suggested in cattle (Simpson et al., 1997).

The present study clearly demonstrated that there is no more 20% of the Shami does resume their cycling activities in August, the early period of breeding season and serum p4 is a reliable and convenient means of monitoring ovarian activity in Shami goats, like in other goat breeds (Shami goats, Zarkawi and Soukouti, 2001; Alpine goat, Thibier et al., 1981; Dwarf goats, Khanum, et al., 2008).

The P4 concentrations at the luteal phase was 1 to 4.5 ng/ml which is comparable to 2.33- 2.94 ng/ml in Dwarf goats (Khanum, et al. (2008) in Pakistan, to 2.6 to 5.4 ng/ml (Khadeja et al., 2005) in Shami goats in Sudan and 1.025 to 8.8 ng/ml (Zarkawi and Soukouti, 2001) in Damascus goats in Syria. However, these variations might be due to genetic, time of breeding season, number of corpora lutea (Jones and Knifton, 1972) and /or body condition (Cortés et al., 2009).

Based on the data presented in this study, the maximal serum IGF-1 concentration (130 ng/ml) preceded the transition elevation in p4 concentration ≥ 1 ng/ml by 3 to 4 days is a good evidence of resumption of cyclic activity occurring estrus and ovulation in Shami goats. This is in agreement with several investigations confirmed that plasma IGF-1 levels increase during the estrus cycle in sheep (Leeuwenberg et al., 1996; Spicer and Zavy, 1992) and goats (Hashizume et al., 2000). In Shipa goats, plasma IGF-1 increases 2 days prior to estrus and reaches its maximal level during the preovulatory LH surge and is coincident with estrus behavior, before decreasing around ovulation time (Hashizume et al., 2000). In agreement with these data, an increase in IGF-1 concentrations in blood serum of pluriparous goats and the tendency to increase in Boer goats during the first ovarian activity after kidding were observed (Djuricic et al., 2011). Maximal IGF-1 concentration presented in this study in serum of Shami goats preceded the maximal p4 concentration by approximately 12 days (Fig.1) and by 8 days in Boer goats (Djuricic et al., 2011). These observations are in agreement with the systemic and local role of IGF-1 in ovarian cycle. Todd et al. (2010) approved that, in estrogen- positive feedback conditions, IGF-1 signaling in the hypothalamus is necessary for secretion of LH surge. Estrogens increase the IGF-1 mRNA in the uterus (Wathes et al., 1998; Carlsson et al., 1991) and the expression in the uterus is independent of GH but, dependent on estrogen (Jones and Clemmons, 1995; Norstedt et al., 1989) and the uterus is the largest of the female reproductive organs. In which the estrogen receptor is present. *In vitro* it was confirmed that IGF-1 also stimulates angiogenesis in the early corpus luteum (Schams et al., 2002). Moreover, the highest mRNA expression for IGF-1 was observed in the early luteal phase and decreased during the middle and late luteal phase (Schams, 1987). Hence, it was postulated that

the IGF-1 system is essential for the development of the early corpus luteum through the stimulation of angiogenesis, luteinization of granulosa-lutein cells and of progesterone production (Berisha and Schams, 2005).

In this study, the presence of non-significant differences in the IGF-1 concentrations between non-cycling Shami goats and significant differences with a long plateau seen especially during the luteal phase in cycling does might be due to the body weights and considerable individual variation in the level of energetic body reserve (Rondina et al., 2005) specially these animals were raised in conditions with major fluctuations in feed availability throughout of the year which can result in the quantity and quality of food intake and, therefore affect body condition. Feed intake influences IGF-1 synthesis in the liver, playing an important role as a metabolic signal in the regulation of the first ovulation (Braw-tal et al., 2004) and the influence of poor energetic status on cyclic activity has been well established (Spicer et al., 1990; Lucy, 2000).

CONCLUSIONS

In conclusion, it was established that serum IGF-1 concentrations in serum of Shami goats follow the cycling activity during the early breeding season period and it is correlated negatively and significantly with the p4 concentrations. The peak of IGF-1 preceded the p4 peak by 12 days in cycling does. Determination the concentrations of these two hormones can help in assessing the reproductive status of Shami goats and facilitate the ability of producers to select females most suited for particular nutritional environments and higher reproductive efficiency.

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REFERENCES

- Abdullah, R., Salleh, M.S., Wan Khadijah ,W.E., Mavrogenis, A.P., 2012. Reproductive, Production and Economic Performances of the Damascus (Shami) Goat of Cyprus. Proceedings of the First Asia Dairy Goat Conference, April 9-12, 2012, Corus Hotel Kuala Lumpur, Malaysia, 106-107.

- Alkhouri, F., 1996. Encyclopedia of Goat Breeds In Arab Countries, ACSAD/AW/N/201, Damascus, Syria.
- Armstrong, DG., Webb, R., 1997. Ovarian follicular dominance: the role of intraovarian growth factors and novel proteins. *Reviews of Reproduction*. 2, 139-146.
- Armstrong, DG., Baxter, G., Gutierrez, CG., Hogg, CO., Glazyrin, AL., Campbell, BK., Bramley, TA., Webb, R., 1998. Insulin-like growth factor binding protein-2 and -4 messenger ribonucleic acid expression in bovine ovarian follicles: effect of gonadotropins and developmental status. *Endocrinology*. 139, 2146-2154.
- Bauernfeind, M., Holtz, W., 1991. Progesterone and estrogen levels in serum of cycling goats measured by enzyme immunoassay. *Small Ruminant Research*. 6, 95-102.
- Berisha, B., Schams, D., 2005. Ovarian function in ruminants. *Domestic Animal Endocrinology*. 29, 305-317.
- Braw-Tal, R., Kaim, M., Pen, S., Lavon, Y., 2004. First post partum ovulation in high yielding dairy cows: association with body condition, milk production and plasma IGF-1 level. *Reproduction in Domestic Animals*. 39, 278-279.
- Carlsson, B., Billing, G., 1991. Insulin-like growth factor-1 gene expression during development and estrus cycle in the rat uterus. *Molecular Cell Endocrinology*. 77, 175-180.
- Carlsson, B., Carlsson, L., Billig, H., 1989. Estrus cycle-dependent covariation of insulin-like growth factor -1 (IGF-1) messenger ribonucleic acid and protein in the rat ovary. *Molecular Cell Endocrinology*. 64, 271-276.
- Constantinou, A., 1989. Genetic and environmental relationship, of body weight, milk yield and litter size in Damascus goats. *Small Ruminant Research*. 2, 163-174.
- Cortés, E., Vera-Avila, H.R., Urrutia-Morales, J., Villagómez-Amezcuca, E., Jiménez-Severiano, H., Mejía-Guadarrama, C.A., Rivera-Lozano, M.T., GámezVázquez, H.G., 2009. Nutritional status influences reproductive seasonality in Creole goats: 1. Ovarian activity during seasonal reproductive transitions. *Animal Reproduction Science*. 116, 282-290
- Delgadillo, J.A., Fitz-Rodríguez, G., Duarte, G., Veliz, F.G., Carrillo, E., Flores, J.A., Vielma, J., Hernandez, H. and Malpaux, B., 2004. Management of photoperiod to control caprine reproduction in the subtropics. *Reproduction Fertility and Development*. 16, 471-478.
- Demeestere, I., Gervy, C., Centner, J., Devreker, F., Englert, Y., Delbaere, A., 2004. Effect of insulin-like growth factor-I during preantral follicular culture on steroidogenesis, *in vitro* oocyte maturation, and embryo development in mice. *Biology of Reproduction*. 70, 1664-1669.

- Djuricic, D., Filipovic, N., Doranic, T., Lipar, M., Prvanovic, N., Turk, R., Gracner, D., Stanin, D., Folinozic, I., Samardzija, M., 2011. Progesterone and insulin-like growth factor I levels in blood of Boer goats during puerperium out-of-season in a mild climate region. *Reproduction in Domestic Animals*. 46, 776-780.
- El-Hommosy, F.S., Salem, I.A., Allam, S., Salem, A., 1991. Ovarian hormones throughout the estrous cycle in goat under upper Egypt conditions' *Assiut Veterinary Medicine*. 25, 46-55.
- Forcada, F., Abecia, J.A., 2006. The effect of nutrition on the seasonality of reproduction in ewes. *Reproduction Nutrition Development*. 46, 355-365.
- Geisert, R.D., Lee, C. Y. Simmen, F.A., Zavy, M.T., Fliss, A.E., Bazer, F.W., Simmen, R.C.M., 1991. Expression of messenger RNAs encoding insulin-like growth factor-I and -II and insulin-like growth factor binding protein-2 in bovine endometrium during the estrous cycle and early pregnancy. *Biology of Reproduction*. 45, 975-983.
- Giudice, L.C., 1992. Insulin-like growth factors and ovarian development. *Endocrine Reviews*. 13, 641-669
- Gutierrez, C.G., Ralph, J.H., Telfer, E. E., Wilmut, I., Webb, R., 2000. Growth and antrum formation of bovine antral follicles in long-term culture *in vitro*. *Biology of Reproduction*. 62, 1322-1328.
- Hashizume, T., K. Ohtsuki and N. Matsumoto. 2000. Serum insulin-like growth factor-I concentrations increase during the estrous phase in goats. *Domestic Animal Endocrinology*. 18, 253-263.
- Hossner, K.L., McCusker, R.H., Dodson, M.V., 1997. Insulin-like growth factors and their binding proteins in domestic animals. *Animal Science*. 64, 1-15.
- Hunter, M.G., Robinson, R.S., Mann, G.E., Webb, R., 2004. Endocrine and paracrine control of follicular development and ovulation rate in farm. *Animal Reproduction Science*. 82-83, 461-467.
- Jones, J.I., Clemmons, D.R., 1995. Insulin-like growth factors and their binding proteins: biological actions, *Endocrine Review*. 16, 3-34.
- Jones, D.E., Knifton, A., 1972. Progesterone concentration in the peripheral plasma off goat during the oestrus cycle. *Research in Veterinary Science*. 13, 193-195.
- Khadiga, M., Gaafar, M.K., Doaa, G.,Teleb, F., 2005. The hormonal profile during the estrous cycle and gestation in Damascus goats. *Small Ruminant Research*. 57, 85-93.
- Khanum, S.A., Hussain, H., Kausar, R., 2008. Progesterone and estradiol profile during estrus cycle and gestation in dwarf goats (*Capra Hircus*). *Pakistan Veterinary Journal*. 28, 1-4.
- Ko, Y., Lee, C.Y., Ott, T.L., Davis, M.A., Simmen, R.C.M., Bazer, F.W., Simmen, F.A., 1991. Insulin-like growth factors in sheep uterine fluids:

- concentrations and relationship to ovine trophoblast protein-1 production during early pregnancy, *Biology of Reproduction*. 45, 135-142.
- Lalman, D. L., J. E. Williams, B. W. Hess, M. G. Thomas and D. H. Keisler. 2000. Effect of dietary energy on milk production and metabolic hormones in thin primiparous beef heifers. *J. Anim. Sci.* 78, 530-538.
- Leeuwenberg, B.R., Hurst, P.R., McNatty, K.P., 1995. Expression of IGF-I mRNA in the ovine ovary. *Journal of Molecular Endocrinology*. 15, 251-258.
- Littell, R.C., Henry P.R., Ammerman C.B., 1998. Statistical analysis of repeated measures data using SAS procedures. *Journal of Animal Science*. 76, 1216-1231.
- Louhio, H., Hovatta, O., Sjoberg, J., Tuuri, T., 2000. The effects of insulin, and insulin-like growth factors I and II on human ovarian follicles in long-term culture. *Molecular Human Reproduction*. 6, 694-698.
- Lucy, M.C., 2000. Regulation of ovarian follicular growth in somatotropin and insulin - like growth factor -1 in cattle. *Journal of Dairy Science*. 83, 1635- 1647.
- Mani, A.U., McKelvey, W.A.C., Watson, E.D., 1996. Effect of undernutrition on gonadotropin profiles in nonpregnant, cycling goats. *Animal Reproduction Science* 43, 25-33.
- Mani, A.U., McKelvey, W.A., Watson, E.D., 1992. The effects of low level of feeding on response to synchronization of estrous, ovulation rate and embryo loss in goats. *Theriogenology*. 36, 1013-1022.
- Martin, G.B., Tjondronegoro, S., Boukhliq, R., Blackberry, M.A., Briegel, J.R., Blache, D., Fisher, J.A., Adams, N.R., 1999. Determinants of the annual pattern of reproduction in mature male Merino and Suffolk sheep: modification of endogenous rhythms by photoperiod. *Reproduction Fertility and Development*. 11, 355-366.
- Mavrogenis, A.P., 1988. Control of the reproductive performance of Chios sheep and Damascus goats. *Proceedings of the Final Research Coordination Meeting, Rabat, March 23-27, 1987, International Atomic Energy Agency, Vienna, Austria*, 151-172.
- Mavrogenis, A.P., Antoniadis, N.Y., Hooper, R.W., 2006. The Damascus (Shami) goat of Cyprus, *AGRI*. 38, 57-65.
- Millard, V., Uzberkova, S., Guignot, F., Perreau, C., Rame, C., Curyal-Castel, S., anddupont, J., 2010. Effect of adenopectin on bovine granulosa cell steroidogenesis, oocyte maturation and embryo development. *Reproduction Biology and Endocrinology*. 8, 23-32.
- Morrison CD, Wood R, McFadin-Buff EL, Whitley NC, Keisler DH. 2002. Effect of intravenous infusion of recombinant ovine leptin on food intake and serum concentrations of GH, LH, insulin, IGF-I, cortisol, and thyroxine in growing prepuberal ewe lambs. *Dom Anim Endo* 22,103-

112.

- Nonaka, S., Tsutomu Hashizume, T., Horiuchi, M., Utako Mikami, U., Osawa, T., Miyake, Y., Hara, S., 2003. Origin of plasma insulin-like growth factor-I (IGF-I) during estrus in goats. *Journal of Reproduction and Development*. 49, 253-258.
- Norstedt, G., Levinovitz, A., Eriksson, H., 1989. Regulation of uterine insulin-like growth factor I mRNA and insulin-like growth factor II mRNA by estrogen in the rat. *Acta Endocrinologica Copenhagen*. 120, 466-472.
- Paula, N.R.O., Galeati, G., Teixeira, D.I.A., Lopes, Junior, E.S., Freitas, V.J.F., Rondina, D., 2005. Responsiveness to progestagen-eCG-cloprostenol treatment in goat food restricted for long period and refed. *Reproduction in Domestic Animals* 40, 108-110.
- Pathiraja, N., Oyedipe, E.O., Gyang, E.O., Obasi, A., 1991. Plasma progesterone levels during oestrous cycle and their relationship with the ovulation rate in Red Sokoto (Maradi) goats. *British Veterinary Journal*. 147, 57-62.
- Perks, C.M., Dennining-Kendall, P.A., Gilmour, R.S., Wathes, D.C., 1995. Localization of messenger ribonucleic acids for insulin-like growth factor I (IGF-I), IGF-II and the type 1 IGF receptor in the ovine ovary throughout the estrous cycle. *Endocrinology*. 136, 5266-5273.
- Pohler KG, Pereira MHC, Lopes FR, Lawrence JC, Keisler DH, Smith MF, Vasconcelos JLM, Green JA. 2015. Circulating concentrations of bovine pregnancy associated glycoproteins and late embryonic mortality in lactating dairy herds. *J. Dairy Science*. 99,1-11.
- Richards, M.W., Wettemann, R.P., Spicer, L.J., Morgan, G.L., 1991. Nutritional anestrus in beef cows: Effect of body condition and ovariectomy on serum luteinizing hormone and insulin - like growth factor-1. *Biology of Reproduction*. 44, 961-966.
- Rondina, D., Freitas, V.J.F., Spinaci, M., Galeati, G., 2005. Effect of nutrition on plasma progesterone levels, metabolic parameters and small follicles development in unstimulated goats reared under constant photoperiod regimen. *Reproduction in Domestic Animals*. 40, 548-552.
- Schams, D., 1987. Luteal peptides and intercellular communications. *Journal of Reproduction and Fertility*. Supplement, 34, 87-99.
- Schams, D., Bensha, B., Krosmann, M., Amselgruber, W.M., 2002. Expression and localization of IGF family members in bovine antral follicles during final growth and in luteal tissue during different stages of estrous cycle and pregnancy. *Domestic Animal Endocrinology*. 22, 51-72.
- Schmidt, A., Einspanier, R., Amselgruber, W., Sinowatz, F., Schams, D., 1994. Expression of insulin-like growth factor 1 (IGF-1) in the bovine oviduct during the oestrous cycle. *Experimental Clinical Endocrinology*. 102, 364-369

- Schneider, J.E., 2004. Energy balance and reproduction. *Physiology of Behavior*. 81, 289–317.
- Shalaby, A.S., Sharawy, S.M., Saleh, N.H., Medan, M.S., 2002. Productive performance of Damascus goats in semi-arid areas in Egypt. In: *Proceedings of the 7th International Conference on Goats, France, May 15–21*, pp: 424–425.
- Shelton, M., 1991. Management of reproduction in the goat, VII Reunion Nationals Obre Caprinocultura, Monterrey, N.L. Mexico, 168-184.
- Simpson, R.B., Chase, C.C., Spicer, L.J., Carroll, J.A., Hammond, A.C., Welsh, T.H., 1997. Effect of exogenous estradiol on plasma concentrations of somatotropin, insulin-like growth factor binding protein activity, and metabolites in ovariectomized Angus and Brahma cows. *Domestic Animal Endocrinology*. 14, 367- 380.
- Spicer, L.I., Tucher W.B., Adams, G.D., 1990. Insulin- like growth factor -1 in dairy cows: relationship among energy balance, body condition, ovarian activity and estrus behavior. *Journal of Dairy Science*. 73, 929-937.
- Spicer, L.J., Alpizar, E., Echterkamp, S.E., 1993. Effects of insulin, insulin-like growth factor I, and gonadotropins on bovine granulosa cell proliferation, progesterone production, estradiol production, and (or) insulin-like growth factor I production *in vitro*. *Journal of Animal Science*, 71, 1232–1241.
- Spicer, L.J., Zavy, M.T., 1992. Concentrations of insulin-like growth factor-I in serum of sheep with different ovulation rates: changes during the estrous cycle. *Theriogenology*. 37, 395–405.
- Stevenson, K.R., Wathes, D.C., 1996. Insulin-like growth factors and their binding proteins in the ovine oviduct during the oestrous cycle. *Journal of Reproduction and Fertility*. 108, 31–40.
- Stevenson, K.R., Gilmour, R.S., Wathes, D.C., 1994. Localization of insulin-like growth factor-I (IGF-I) and -II messenger ribonucleic acid and type 1 IGF receptors in the ovine uterus during the estrous cycle and early pregnancy. *Endocrinology*. 134, 1655–1664.
- Tamadon, A., Kafi, M., Saeb, M., Mirzaei, A., Saeb, S., 2010. Relationship between insulin-like growth factor -1 milk yield, body condition score and postpartum luteal activity in high- producing dairy cows. *Tropical Animal Health and Production*. 43, 29-34.
- Tanaka, T, Fujiwara, KI, Kim, S, Kamomae, H., Kaneda Y., 2004. Ovarian and hormonal responses to a progesterone releasing controlled internal drug releasing treatment in dietary-restricted goats. *Animal Reproduction Science*. 84, 135–146.
- Thibier, M., Pothélet, D., Jeanguyot, N., De Montigny, G., 1981. Estrus behavior, progesterone in peripheral plasma and milk in dairy goats at onset of breeding season. *Journal of Dairy Science*. 64, 513-519.

- Todd, B.J., MerhiZo, Shu, J., Etgen, A.M., Neal- Perry, G.S., 2010. Hypothamic insulin-like growth factor-1 receptors are necessary for hormone-dependent luteinizing hormone surges: Implications for female reproductive aging. *Endocrinology*. 151, 1356-366.
- Yu, Y., Li, W., Han, Z., Luo, M., Chang, Z. Tan, J., 2003. The effect of follicle stimulating hormone on follicular development, granulosa cell apoptosis and steroidogenesis and its mediation by insulin-like growth factor -1 in the goat ovary. *Theriogenology*. 60, 1691-1704.
- Wathes, D.C., Reynolds, T.S., Robinson, R.S., Stevenson, K.R., 1998. Role of the insulin-like growth factor system in uterine function and placental development in ruminants. *Journal of Dairy Science*. 81, 1778-89.
- Woad, K.J., Baxter, G., Hogg, C.O., Bramley, T.A., Webb, R., Armstrong, D.G., 2000. Expression of mRNA encoding insulin-like growth factors I and II and type 1 IGF receptor in the bovine corpus luteum at defined stages of the oestrous cycle. *Journal of Reproduction and Fertility*. 120, 293-302.
- Zarkawi, M., 2007. Oestrous synchronisation and fertility in cycling Damascus does using the synthetic prostaglandin F₂ α , Iliren. *Journal of Applied Animal Research*. 32, 37- 40.
- Zarkawi, M., Al-Masri, M. R., 2002. Use of radioimmunoassay to measure progesterone levels during different reproductive stages in female Damascus goats. *Tropical Animal Health and Production*. 34, 535-539.
- Zarkawi, M., Al-Merestani, M.R., Wardeh, M.F., 2003. Serum progesterone levels for diagnosing pregnancy and monitoring corpora lutea function during different reproductive stages in hormonally-treated female Damascus goats. Sixth Arab Conference on the Peaceful Uses of Atomic Energy, Cairo, Egypt, 14-18 December 2002, Volume 3, pp. 443-456.
- Zarkawi, M., Soukouti, A., 2001. Serum progesterone levels using radioimmunoassay during oestrous cycle of indigenous Damascus does. *New Zealand Journal of Agricultural Research*. 44, 165-169.
- Zhou, J., Wang, J., Penny, D, Monget, P., Arraztoa, J.A., Fogelson, L.J., Bndy, C.A., 2003. Insulin-like growth factor binding protein 4 expression parallels luteinizing hormone receptor expression and follicular luteinization in the primate ovary. *Biology of Reproduction*. 69, 22-29.